

Original Article

Enhancing Radiosensitivity in Prostate Adenocarcinoma by Geranyloxy coumarin: The Potential Roles of β -Catenin, c-MYC, and PSMD10

Nafiseh Bagheri, Yasaman Abolhassani, Fatemeh B. Rassouli, Khadijeh Jamialahmadi

DOI: 10.34172/PS.026.43308

To appear in: Pharmaceutical Science (<https://pstbzmed.com/>)

Received date: 29 Sep 2025

Revised date: 1 Feb 2026

Accepted date: 21 Feb 2026

Please cite this article as: Bagheri N, Abolhassani Y, Rassouli FB, Jamialahmadi K. Enhancing radiosensitivity in prostate adenocarcinoma by geranyloxy coumarin: The potential roles of β -catenin, c-MYC, and PSMD10 . Pharm Sci. 2026. doi: 10.34172/PS.026.43308

This is a PDF file of a manuscript that have been accepted for publication. It is assigned to an issue after technical editing, formatting for publication and author proofing.

Enhancing Radiosensitivity in Prostate Adenocarcinoma by Geranyloxy coumarin: The potential Roles of β -Catenin, c-MYC, and PSMD10

Nafiseh Bagheri ^{a,1}, Yasaman Abolhassani ^{a,1}, Fatemeh B. Rassouli ^{b*}, Khadijeh Jamialahmadi ^{c*}

^a Department of Medical Biotechnology and Nanotechnology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.

^b Novel Diagnostics and Therapeutics Research Group, Institute of Biotechnology, Ferdowsi University of Mashhad, Mashhad, Iran

^c Biotechnology Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran.

¹These authors contributed equally to this work as the first authors.

Running title: Radiosensitizing Role of Geranyloxy coumarin in Prostate Cancer

* Corresponding Authors:

Dr. Khadijeh Jamialahmadi

Professor of Pharmaceutical Biotechnology, Biotechnology Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran.

Email: jamialahmadikh@mums.ac.ir

Tel : +98-513-800-2293

Dr. Fatemeh B. Rassouli

Associate Professor, Novel Diagnostics and Therapeutics Research Group, Institute of Biotechnology, Ferdowsi University of Mashhad, Mashhad, Iran.

Email: behnam3260@um.ac.ir

Tel : +98-513-880-5504

ORCID

Khadijeh Jamialahmadi (<https://orcid.org/0000-0003-2173-003X>)

Fatemeh B. Rassouli (<https://orcid.org/0000-0003-1889-0964>)

Yasaman Abolhassani (<https://orcid.org/0000-0003-4700-5186>)

Nafiseh Bagheri (<https://orcid.org/0009-0000-6719-4645>)

Conflict of interest

The author states no conflict of interest.

Author contributions

Conceptualization: Fatemeh B. Rassouli, Khadijeh Jamialahmadi

Methodology & Formal analysis: Nafiseh Bagheri, Yasaman Abolhassani, Fatemeh B. Rassouli

Investigation: Nafiseh Bagheri, Yasaman Abolhassani

Visualization: Nafiseh Bagheri, Yasaman Abolhassani

Supervision: Fatemeh B. Rassouli, Khadijeh Jamialahmadi

Funding acquisition: Fatemeh B. Rassouli, Khadijeh Jamialahmadi

Writing – original draft: Nafiseh Bagheri, Yasaman Abolhassani

Writing – review & editing: Fatemeh B. Rassouli, Khadijeh Jamialahmadi

Nafiseh Bagheri and Yasaman Abolhassani contributed equally to this work. All authors read and approved the final manuscript.

Funding

This work was supported by the Vice-Chancellor for Research and Technology, Mashhad University of Medical Sciences (Grant No.: 4011854; Ethical code: IR.MUMS.MEDICAL.REC.1401.674) and Ferdowsi University of Mashhad, Mashhad, Iran.

Acknowledgment

This work was supported by the Vice-Chancellor for Research and Technology, Mashhad University of Medical Sciences and Ferdowsi University of Mashhad, Mashhad, Iran.

Abstract

Background: Prostate adenocarcinoma is commonly treated with radiation and chemotherapy, but resistance and toxicity limit their success, highlighting the need for novel radiosensitizers.

We investigated the effects of 7-Geranyloxy coumarin, alone and in combination with radiation, on the expression of β -catenin (*CTNNB1*), *c-MYC*, and Gankyrin (*PSMD10*), key mediators of Wnt signaling pathway associated with radioresistance in prostate adenocarcinoma cells.

Methods: STRING and GEPIA2 were used for interactome mapping, pathway enrichment, and expression/survival validation in prostate adenocarcinoma. Human prostate adenocarcinoma (PC-3) cells were pretreated with 40 μ M 7-Geranyloxy coumarin and subsequently irradiated with 4 Gy. Gene expression was assessed by real-time PCR after 72 h.

Results: In silico analyses confirmed interactions among *CTNNB1*, *c-MYC*, and *PSMD10*, their involvement in the Wnt pathway, and the overexpression and prognostic trends of *c-MYC* and *PSMD10*. In PC-3 cells, 7-Geranyloxy coumarin treatment significantly decreased *c-MYC* expression ($p < 0.0001$), while the combined treatment increased *CTNNB1* ($p < 0.01$) but decreased *PSMD10* and *c-MYC*, indicating disruption of Wnt pathway.

Conclusion: 7-Geranyloxy coumarin modulates the Wnt signaling pathway by suppressing the expression of *CTNNB1*, *c-MYC*, and *PSMD10*, key mediators of this pathway, thereby enhancing radiosensitivity and highlighting its potential as an adjuvant therapy in prostate adenocarcinoma.

Keywords: 7-Geranyloxy coumarin, Prostate adenocarcinoma, Radiation therapy, Gankyrin, β -catenin, *c-MYC*.

Introduction

Prostate adenocarcinoma is the second most prevalent cancer in men worldwide,^{1,2} and it has just been identified as the third most prevalent cancer in Iranian men.^{3,4} The treatment of prostate cancer includes surgery, chemotherapy, radiotherapy, hormone therapy and their combination.⁵⁻⁸ Simultaneous chemoradiotherapy is superior to radiotherapy alone or to alternating radiotherapy and chemotherapy.^{9,10}

Natural products represent a promising frontier in cancer therapy, enhancing tumor cell sensitivity to radiation and significantly improving the outcomes of simultaneous chemoradiotherapy.¹⁰⁻¹² Radiosensitizers increase the sensitivity of tumor cells to radiation, allowing for lower doses while maintaining therapeutic efficacy.^{13,14}

7- Geranyloxy coumarin (also known as auraptene) is a natural coumarin derivative found in citrus plant extracts that can be synthesized under laboratory conditions. This coumarin has various biological activities, such as antibacterial, antifungal, anti-inflammatory, and antioxidant properties, and its anti-cancer and cancer-prevention effects have been demonstrated in numerous studies.¹⁵⁻¹⁹ Its synergistic effects with chemotherapy drugs have also been reported in skin, esophageal, and colon cancer cells.^{9,20,21}

The Wnt/ β -catenin pathway, a key regulator of proliferation, differentiation, and metastasis, can be modulated by coumarins.²² Wnt pathway genes can drive radioresistance by modulating other genes or pathways.²³ For example, β -catenin regulates LIG4 expression, while WISP1 directly phosphorylates histone 2 (γ -H2AX). The β -catenin/TCF transcriptional complex also regulates the expression of ALDH. Together, LIG4, γ -H2AX, and ALDH enhance DNA damage repair capacity, contributing to radioresistance.^{24,25}

c-MYC is one of the most frequently activated oncogenes, estimated to be involved in ~20% of all human cancers. As a transcription factor, it regulates genes controlling proliferation,

apoptosis, and DNA repair.²⁶ In breast cancer, c-MYC activates Wnt signaling by suppressing the inhibitors DKK1 and SFRP1, although the detailed mechanisms and biological significance remain unclear.²⁷

PSMD10 (Gankyrin), a proteasome assembly chaperone, is an oncoprotein that is upregulated in a variety of cancers.²⁸ Gankyrin also promotes Wnt/ β -catenin signaling by stabilizing and enhancing both the β -catenin protein expression and transcriptional activity which drives transcription of Wnt target genes like c-MYC. In turn, β -catenin and c-MYC can transcriptionally upregulate *PSMD10*, forming a positive feedback loop that amplifies Wnt pathway activation and tumor progression.^{28,29} Building on our previous study that demonstrated the radiosensitizing effects of 7-Geranyloxy coumarin in prostate adenocarcinoma cells, we aimed to investigate the underlying molecular mechanisms. To our knowledge, no previous study has examined how 7-Geranyloxy coumarin modulates key Wnt/ β -catenin signaling components (*CTNNB1*, *c-MYC*, and *PSMD10*) in prostate adenocarcinoma. Therefore, we analyzed the effects of 7-Geranyloxy coumarin, alone and in combination with radiation, on their expression in PC-3 cells, a cell line derived from androgen-independent metastatic prostate adenocarcinoma. This model replicates aggressive, radioresistant disease and exhibits high Wnt activity, making it an ideal model for evaluating radiosensitizers using real-time PCR.

Material and Methods

Computational analyses

For interactome mapping of *CTNNB1*, *c-MYC*, and *PSMD10*, the STRING database (<https://string-db.org/>) was used, and gene set enrichment analysis was performed using Reactome pathways, with results reported as false discovery rate (FDR). To validate the expression of *CTNNB1*, *c-MYC*, and *PSMD10* in prostate adenocarcinoma samples compared

to normal specimens, GEPIA 2.0 (<http://gepia2.cancer-pku.cn/>) was utilized; this tool leverages data from the TCGA and GTEx databases.

Survival analysis was conducted using GEPIA2 with default settings (median cutoff for high/low expression groups) on TCGA-PRAD data. STRING was selected for its comprehensive protein-protein interaction database, integrating experimental and predicted interactions. GEPIA2 was chosen for its seamless integration of TCGA and GTEx data, enabling robust expression and survival comparisons.

In vitro gene expression analysis

To determine the effects of 7-Geranyloxy coumarin, both alone and in combination with radiation, on PC-3 cells, RNA samples previously generated and reported in our earlier study Abolhassani et al., 2023 were used. Briefly, PC-3 cells were pretreated with 40 μ M 7-Geranyloxy coumarin and irradiated with 4 Gy X-ray (Elekta Compact™ linear accelerator, Crawley). After 72 h recovery, total RNA was extracted from treated cells and their relevant controls using the RiboEx Total RNA kit (GeneAll). The extracted RNAs were eluted using diethyl pyrocarbonate-treated water. The concentration and purity (260/280 and 260/230 ratios) were measured in duplicate using a NanoDrop™ 2000c spectrophotometer (Thermo Scientific). To confirm RNA integrity, aliquots of RNA samples were electrophoresed on 1.5% agarose gel. Bands of 28S, 18S, and 5S rRNAs were observed, indicating RNA integrity. Reverse transcription of total RNA to cDNA was then performed according to the manufacturer's protocol (Yekta Tajhiz Azma). Using specific primers listed in Table 1, real-time PCR was performed with SYBR Green master mix (Pars Tous) following the manufacturer's instructions. The PCR cycling conditions were as follows: 95 °C for 5 min to activate the polymerase and perform initial denaturation, followed by 40 cycles of 95 °C for 5 s and 60 °C for 30 s. The

melting curves of the PCR products were monitored at the end of each reaction to evaluate PCR specificity. Gene expression was normalized to GAPDH using the $2^{-\Delta\Delta Ct}$ method.

Table 1. The primer sequences used for real-time PCR.

Gene name	Amplicon length (bp)	Primer sequence (5' → 3')
GAPDH	205	Forward: 5'- GGA TTT GGT CGT ATT GGG-3' Reverse: 5'- GGA AGA TGG TGA TGG GATT-3'
<i>c-MYC</i>	159	Forward: 5'- ACTCTGAGGAGGAACAAGAA -3' Reverse: 5'- TGGAGACGTGGCACCTCTT -3'
<i>CTNNB1</i>	76	Forward: 5'- GCTTTCAGTTGAGCTGACCA -3' Reverse: 5'- AAGTCCAAGATCAGCAGTCTCA -3'
<i>PSMD10</i>	155	Forward: 5'- AGCAGCCAAGGGTAACTTGA -3' Reverse: 5'- TACTTGCTCCTTGGGACACC -3'

Statistical analysis

The results were statistically analyzed using GraphPad Prism version 9.0. One-way ANOVA tests followed by Tukey's post-hoc test was used for multiple comparisons. Real-time PCR experiments were conducted in triplicate across three independent experiments. The data are presented as mean \pm SEM, and statistically significant differences were defined as a p-value less than 0.05.

Results

In Silico Analysis of CTNNB1, c-MYC, and PSMD10 in Prostate Cancer

In silico analysis using STRING revealed an interaction network among *CTNNB1*, *c-MYC*, and *PSMD10*, consisting of 23 nodes and 79 edges, with a highly significant enrichment score ($p < 1.0e-16$) (Figure 1A). Reactome pathway enrichment identified several significant terms, including “Deactivation of the β -catenin transactivating complex” (FDR = $1.65e-17$) and “TCF-dependent signaling in response to WNT” (FDR = $2.38e-17$) (Figure 1B). Moreover, validation of expression profiles using GEPIA2 showed that *c-MYC* and *PSMD10* were significantly overexpressed in prostate adenocarcinoma tissues ($n = 492$) compared with normal samples ($n = 154$).

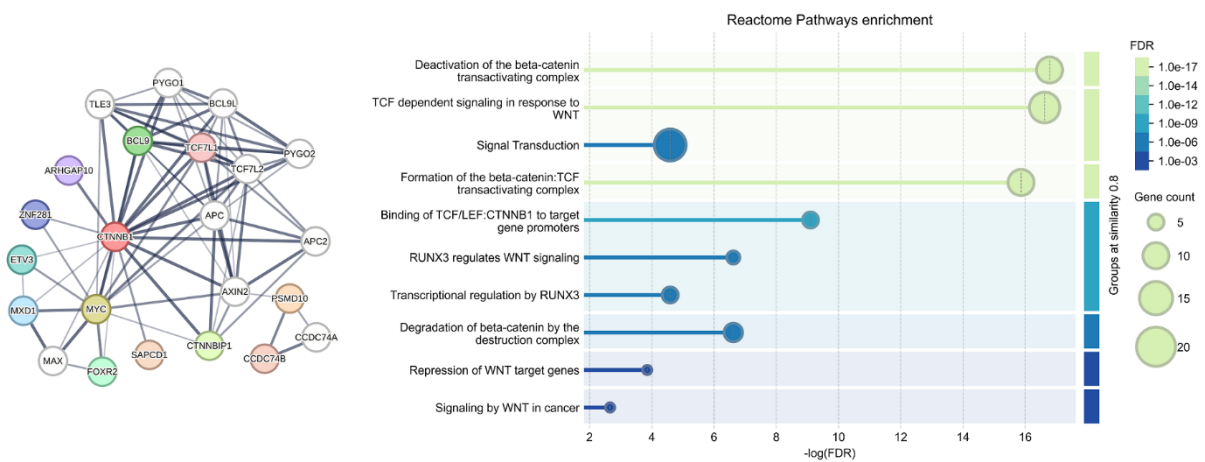


Figure 1. In silico interactome and pathway enrichment analysis. (A) Protein–protein interaction networks for *CTNNB1*, *c-MYC*, and *PSMD10* and their related partners were constructed using STRING. (B) Reactome pathway enrichment analysis highlighting significant terms, including “Deactivation of the β -catenin transactivating complex” and “TCF-dependent signaling in response to WNT.”

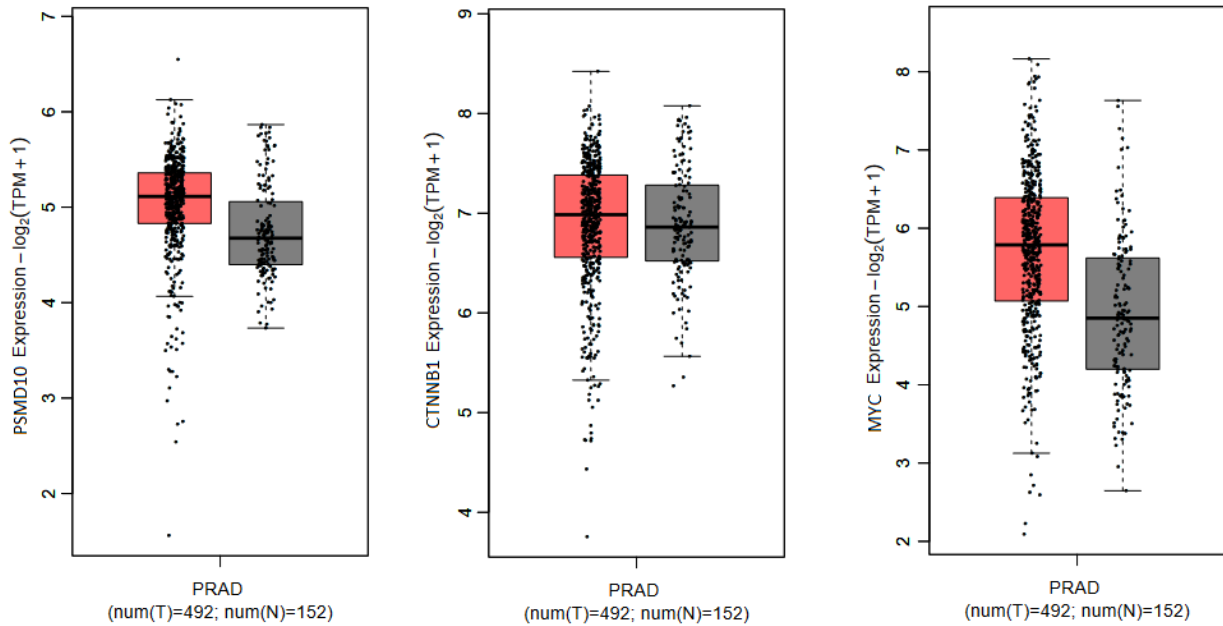


Figure 2. Validation of *CTNNB1*, *c-MYC*, and *PSMD10* expression in prostate adenocarcinoma using GEPIA2. Box plots represent transcript levels in tumor tissues (n = 492) compared with normal prostate samples (n = 154). Results indicate significant overexpression of *c-MYC* and *PSMD10* in tumor tissues, while *CTNNB1* showed no marked difference between the two groups. Data are presented as mean \pm SEM.

To further validate the clinical relevance, survival correlations were assessed using TCGA prostate adenocarcinoma data via GEPIA2 (Figure 3). High *c-MYC* expression was associated with poorer overall survival (HR=1.902, 95% CI: 0.962-3.76, p=0.0645), whereas *PSMD10* showed a trend toward a worse prognosis (HR=6.265, 95% CI: 0.735-53.409, p=0.0933). In contrast, *CTNNB1* expression exhibited a protective trend (HR=0.257, 95% CI: 0.059-1.121, p=0.0707) (Table 2). These findings support the oncogenic roles of *c-MYC* and *PSMD10* in prostate adenocarcinoma progression.

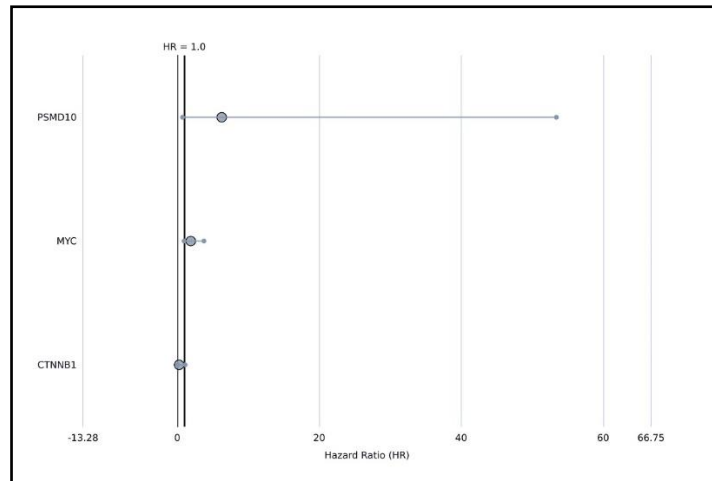


Figure 3. Survival correlations of *CTNNB1*, *c-MYC*, and *PSMD10* in prostate adenocarcinoma, analyzed using TCGA data via GEPIA2.

Effect of 7-Geranyloxy coumarin on gene expression level

As shown in Figure 4, real-time PCR analysis revealed that treatment with 7-Geranyloxy coumarin significantly reduced *c-MYC* expression in PC-3 cells compared to the control group ($p < 0.0001$). A significant decrease in *c-MYC* expression was also observed following radiation treatment alone ($p < 0.0001$). Similarly, DMSO-treated cells exposed to radiation showed a significant reduction in *c-MYC* expression ($p < 0.001$).

In contrast, combined treatment with 7-Geranyloxy coumarin and radiation significantly increased *CTNNB1* expression compared with the control ($p < 0.01$), whereas no significant changes were observed in the other treatment groups. Moreover, *PSMD10* expression was markedly reduced in both the radiation-only and the combined treatment groups ($p < 0.0001$). This upregulation of *CTNNB1*, although seemingly counterintuitive given its role in Wnt activation, coincided with the downregulation of downstream oncogenes *c-MYC* and *PSMD10*, potentially indicating a disruption of canonical Wnt signaling.

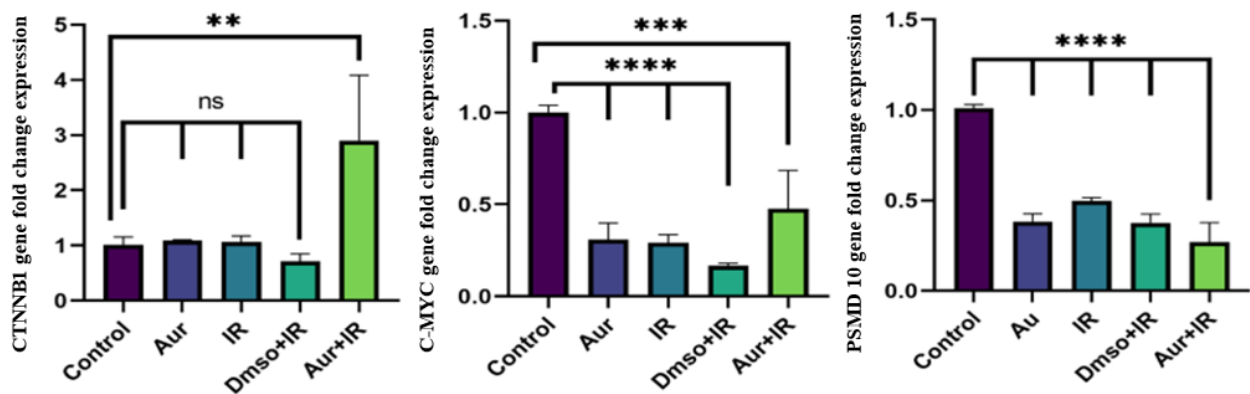


Figure 4. Relative expression of *CTNNB1*, *c-MYC*, and *PSMD10* in PC-3 cells across five treatment groups, as determined by real-time PCR (n=3 biological replicates, each performed in technical triplicates). Gene expression levels were normalized to GAPDH as the internal control. Data are presented as mean \pm SEM. Statistical significance compared with the untreated control group (one-way ANOVA with Tukey's post-hoc test): **p < 0.01 for *CTNNB1* increase in combined treatment; ****p < 0.0001 for *c-MYC* decrease in 7-Geranyloxycoumarin alone, radiation alone, and combined treatment; ***p < 0.001 for *c-MYC* decrease in DMSO-treated irradiated cells; ****p < 0.0001 for *PSMD10* decrease in radiation and combined treatment groups.

Discussion

Prostate adenocarcinoma is one of the most prevalent and lethal cancers affecting men globally, with treatment often involving radiotherapy following chemotherapy or surgery. Since development of radioresistance in cancer cells considerably reduces the clinical response to radiotherapy, there is an urgent need to identify new compounds that enhance the radiosensitivity of prostate adenocarcinoma cells, thereby improving therapeutic outcomes.³⁰⁻³² Previous research has shown that 7-Geranyloxycoumarin has anticancer and antiproliferative properties in various cancer types, including skin, breast, and colon cancers. However, the specific effects of 7-Geranyloxycoumarin on the radiation response in prostate adenocarcinoma cells, as well as the underlying molecular mechanisms, have not been

adequately explored.¹⁵ In our previous study, we investigated the effects of 7-Geranyloxy coumarin on enhancing radiosensitivity of prostate adenocarcinoma cells. The focus of this study was to further elucidate the underlying molecular mechanisms responsible for this effect. Computational analyses in the current study revealed functional and physical associations between *CTNNB1*, *c-MYC*, and *PSMD10* in the protein-protein interaction network. Pathway enrichment analysis further confirmed their involvement in Wnt signaling, a pathway crucial for tumor growth and therapy resistance. Additionally, *c-MYC* and *PSMD10* were upregulated in prostate adenocarcinoma tissue samples. The inclusion of survival correlations further supports the in silico findings, suggesting that *c-MYC* and *PSMD10* may serve as potential prognostic markers, consistent with their overexpression and involvement in radioresistance.

Findings from real-time PCR indicate that in prostate adenocarcinoma cells exposed to radiation, 7-Geranyloxy coumarin changed the expression of *c-MYC*, *CTNNB1* and *PSMD10*. These genes are key components of the Wnt signaling pathway, which regulates cell proliferation, survival, invasion, metastasis, and radiation resistance in prostate adenocarcinoma.^{33,34}

We also found that the expression of *c-MYC* and *PSMD10* genes in PC-3 cells was significantly reduced by 7-Geranyloxy coumarin, both when used alone and in combination with radiation. In contrast, the expression of *CTNNB1* was notably increased. These results suggest that 7-Geranyloxy coumarin enhances the radiosensitivity of prostate adenocarcinoma cells by suppressing the *c-MYC* and *PSMD10* genes, thereby altering Wnt signaling, highlighting its potential as a novel radiosensitizer for prostate cancer therapy.

Oncogenes such as *c-MYC* and *PSMD10* are known to promote cell proliferation, invasion, and metastasis in prostate adenocarcinoma. They also play critical roles in regulating DNA damage

repair, the cell cycle, and apoptosis processes essential for radioresistance. Stabilized *c-MYC* promotes resistance to chemotherapy and radiation by accelerating cell cycle progression and preventing apoptosis. Furthermore, the stabilization and activation of β -catenin have been shown to enhance the growth and survival of prostate adenocarcinoma cells under androgen-deprived conditions. Interestingly, stabilized β -catenin also appears to reduce *c-MYC* activity, a key transcription factor involved in cell growth and proliferation, as well as the motility of prostate adenocarcinoma cells.³⁴⁻³⁶

The PI3K/Akt and Wnt/ β -catenin pathways are both essential for cell survival and radioresistance, and they can be regulated by *PSMD10*.^{24,37} In line with this, our data showed that 7-Geranyloxy coumarin increased *CTNNB1* expression but simultaneously reduced *c-MYC* and *PSMD10*, two oncogenic downstream targets of β -catenin. The observed increase in *CTNNB1* expression following combined treatment appears paradoxical, as elevated β -catenin is typically associated with enhanced radioresistance through Wnt pathway activation.^{23,24} However, this may reflect context-dependent regulation, where β -catenin stabilization could inhibit downstream effectors like *c-MYC* under certain conditions, as seen in androgen-deprived prostate cells.³⁸ In our study, the concurrent downregulation of *c-MYC* and *PSMD10*, both oncogenic targets of β -catenin, supports a potential disruption of the positive feedback loop,^{16,29} contributing to radiosensitization. This hypothesis aligns with our previous functional data showing reduced cell survival,¹⁶ but lacks direct evidence of β -catenin's altered activity.

Rigorous follow-up studies, including β -catenin knockdown experiments, are needed to clarify the molecular mechanisms by which 7-Geranyloxy coumarin regulates β -catenin expression and function in prostate adenocarcinoma cells, and to reconcile these findings with established mechanisms.

Deeper analysis of the Wnt pathway reveals that *PSMD10* enhances β -catenin stability by inhibiting its degradation, while *c-MYC* amplifies the loop through transcriptional activation.^{33,35} Our findings suggest 7-Geranyloxy coumarin may interrupt this at the *PSMD10*- β -catenin interface, possibly via non-canonical mechanisms, as evidenced by *CTNNB1* upregulation without proportional oncogene activation.^{33,35} This could involve crosstalk with PI3K/Akt, where *PSMD10* inhibition reduces Akt-mediated β -catenin phosphorylation.

Our research indicates that 7-Geranyloxy coumarin reduced the expression of *c-MYC* and *PSMD10*, modulating the Wnt signaling pathway and enhancing the radiosensitivity of prostate adenocarcinoma cells. These findings suggest its potential use in combination with radiation therapy to improve clinical outcomes for patients with prostate adenocarcinoma. Future studies should investigate protein expressions, employ pathway inhibitors, and utilize in vivo models to validate radiosensitization and clarify the roles of non-canonical Wnt signaling. Taken together, these results provide a rationale for considering 7-Geranyloxy coumarin as an adjuvant radiosensitizer in prostate adenocarcinoma, which could ultimately improve therapeutic efficacy in clinical practice.

Conclusion

This study demonstrated that 7-Geranyloxy coumarin significantly reduced the expression of *c-MYC* and *PSMD10*, thereby modulating the Wnt signaling pathway and enhancing the radiosensitivity of prostate adenocarcinoma cells. These findings highlight its potential as an adjuvant radiosensitizer in prostate cancer treatment. Although *CTNNB1* upregulation warrants cautious interpretation, the overall effect appears to favor radiosensitization through suppression of downstream oncogenes. Further validation in additional cell lines and preclinical models is warranted to confirm the therapeutic applicability of 7-Geranyloxy coumarin.

Acknowledgments

None

Competing Interests

There is no conflict of interest.

Informed consent

Not applicable,

Use of Large Language Models, AI, and Machine Learning Tools

None declared.

Data availability

Not applicable.

References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2021;71(3):209-49. doi: 10.3322/caac.21660
2. Zhou X, Wei C, Liu X, Zhang Z, Wu Y, Zeng B, et al. Revealing the role of bisphenol a on prostate cancer progression and identifying potential targets: A comprehensive analysis from population cohort to molecular mechanism. *Ecotoxicology and Environmental Safety* 2025;296:118209. doi: 10.1016/j.ecoenv.2025.118209
3. Tayarani-Najaran Z, Tayarani-Najaran N, Eghbali S. A review of auraptene as an anticancer agent. *Front Pharmacol* 2021;12:698352. doi: 10.3389/fphar.2021.698352
4. Wang L, Lu B, He M, Wang Y, Wang Z, Du L. Prostate cancer incidence and mortality: Global status and temporal trends in 89 countries from 2000 to 2019. *Front Public Health* 2022;10:811044. doi: 10.3389/fpubh.2022.811044
5. Li J, He H-G, Guan C, Ding Y, Hu X. Dynamic joint prediction model of severe radiation-induced oral mucositis among nasopharyngeal carcinoma: A prospective longitudinal study. *Radiotherapy and Oncology* 2025:110993. doi: 10.1016/j.radonc.2025.110993
6. Wang Y, Xu Y, Song J, Liu X, Liu S, Yang N, et al. Tumor cell-targeting and tumor microenvironment-responsive nanoplatforms for the multimodal imaging-guided photodynamic/photothermal/chemodynamic treatment of cervical cancer. *International journal of nanomedicine* 2024:5837-58. doi: 10.2147/IJN.S466042
7. Wang ZB, Zhang X, Fang C, Liu XT, Liao QJ, Wu N, et al. Immunotherapy and the ovarian cancer microenvironment: Exploring potential strategies for enhanced treatment efficacy. *Immunology* 2024;173(1):14-32. doi: 10.1111/imm.13793

8. Anjomshoa M, Amirheidari B, Janczak J, Sahihi M, Abolhassani Y, Farsinejad A, et al. In vitro and in silico studies of a zn (ii) complex as a potential therapeutic agent for breast cancer. *Scientific Reports* 2024;14(1):29138. doi: 10.1038/s41598-024-79644-0
9. Saboor-Maleki S, Rassouli FB, Matin MM, Iranshahi M. Auraptene attenuates malignant properties of esophageal stem-like cancer cells. *Technol Cancer Res Treat* 2017;16(4):519-27. doi: 10.1177/1533034616650119
10. Bemidinezhad A, Abolhassani Y, Feiz K, Parsa-Kondelaji M, Soukhtanloo M. Carbon and phosphorus quantum dots: Advancing radiotherapy through innovative radiosensitization. *Biochem Biophys Res Commun* 2025;773:152054. doi: 10.1016/j.bbrc.2025.152054
11. Bemidinezhad A, Abolhassani Y, Sarabian Tabrizi A, Noroozi-Karimabad M, Parsa-Kondelaji M, Roshani R, et al. Aptamers in combination therapies for enhanced radiosensitization in cancer. *Iran J Biotechnol* 2025;23(1):e4032. doi: 10.30498/ijb.2025.491856.4032
12. Mathan SV, Rajput M, Singh RP. Chemotherapy and radiation therapy for cancer. Understanding cancer: Elsevier; 2022. p. 217-36.
13. Bemidinezhad A, Abolhassani Y, Noroozi-Karimabad M, Gholami AA, Alalikhani A, Roshani R, et al. Revolutionizing cancer therapy: Monoclonal antibodies in radiosensitization. *BiolImpacts* 2025;15(1):30996-. doi: 10.34172/bi.30996
14. Chen Y, Deng Y, Li Y, Qin Y, Zhou Z, Yang H, et al. Oxygen-independent radiodynamic therapy: Radiation-boosted chemodynamics for reprogramming the tumor immune environment and enhancing antitumor immune response. *ACS Applied Materials & Interfaces* 2024;16(17):21546-56. doi: 10.1021/acsami.4c00793
15. Hosseinimehr SJ. Beneficial effects of natural products on cells during ionizing radiation. *Rev Environ Health* 2014;29(4):341-53. doi: 10.1515/reveh-2014-0037
16. Abolhassani Y, Mirzaei S, Nejabat M, Talebian S, Gholamhosseinian H, Iranshahi M, et al. 7-geranyloxycoumarin enhances radio sensitivity in human prostate cancer cells. *Mol Biol Rep* 2023;50(7):5709-17. doi: 10.1007/s11033-023-08439-9
17. Jamialahmadi K, Salari S, Alamolhodaei NS, Avan A, Gholami L, Karimi G. Auraptene inhibits migration and invasion of cervical and ovarian cancer cells by repression of matrix metalloproteinases 2 and 9 activity. *J Pharmacopuncture* 2018;21(3):177-84. doi: 10.3831/KPI.2018.21.021
18. Hosseini F, Ahmadi A, Hassanzade H, Gharedaghi S, Rassouli FB, Jamialahmadi K. Inhibition of melanoma cell migration and invasion by natural coumarin auraptene through regulating emt markers and reducing mmp-2 and mmp-9 activity. *Eur J Pharmacol* 2024;971:176517. doi: 10.1016/j.ejphar.2024.176517
19. Kang L, Gao X-H, Liu H-R, Men X, Wu H-N, Cui P-W, et al. Structure–activity relationship investigation of coumarin–chalcone hybrids with diverse side-chains as acetylcholinesterase and butyrylcholinesterase inhibitors. *Molecular diversity* 2018;22(4):893-906. doi: 10.1007/s11030-018-9839-y
20. Genovese S, Epifano F. Auraptene: A natural biologically active compound with multiple targets. *Curr Drug Targets* 2011;12(3):381-6. doi: 10.2174/138945011794815248
21. Kleiner-Hancock HE, Shi R, Remeika A, Robbins D, Prince M, Gill JN, et al. Effects of atra combined with citrus and ginger-derived compounds in human scc xenografts. *BMC cancer* 2010;10(1):394. doi: 10.1186/1471-2407-10-394
22. Pelusi L, Mandatori D, Di Pietrantonio N, Del Pizzo F, Di Tomo P, Di Pietro N, et al. Estrogen receptor 1 (esr1) and the wnt/beta-catenin pathway mediate the effect of the coumarin derivative umbelliferon on bone mineralization. *Nutrients* 2022;14(15):3209. doi: 10.3390/nu14153209

23. Woodward WA, Chen MS, Behbod F, Alfaro MP, Buchholz TA, Rosen JM. Wnt/beta-catenin mediates radiation resistance of mouse mammary progenitor cells. *Proc Natl Acad Sci U S A* 2007;104(2):618-23. doi: 10.1073/pnas.0606599104
24. Yang Y, Zhou H, Zhang G, Xue X. Targeting the canonical wnt/beta-catenin pathway in cancer radioresistance: Updates on the molecular mechanisms. *J Cancer Res Ther* 2019;15(2):272-7. doi: 10.4103/jcrt.JCRT_421_18
25. Klautke G, Muller K. Chemotherapeutic agents for gi tumor chemoradiotherapy overview of chemotherapeutic agents to be combined with radiotherapy in the gi tract and their potential as radiosensitizers. *Best Pract Res Clin Gastroenterol* 2016;30(4):529-35. doi: 10.1016/j.bpg.2016.07.001
26. Dang CV, O'Donnell KA, Zeller KI, Nguyen T, Osthus RC, Li F, editors. The c-myc target gene network. *Seminars in cancer biology*; 2006: Elsevier.
27. Xu J, Chen Y, Huo D, Khramtsov A, Khramtsova G, Zhang C, et al. Beta-catenin regulates c-myc and cdkn1a expression in breast cancer cells. *Mol Carcinog* 2016;55(5):431-9. doi: 10.1002/mc.22292
28. Zamani P, Matbou Riahi M, Momtazi-Borojeni AA, Jamialahmadi K. Gankyrin: A novel promising therapeutic target for hepatocellular carcinoma. *Artif Cells Nanomed Biotechnol* 2018;46(7):1301-13. doi: 10.1080/21691401.2017.1388250
29. He F, Chen H, Yang P, Wu Q, Zhang T, Wang C, et al. Gankyrin sustains pi3k/gsk-3beta/beta-catenin signal activation and promotes colorectal cancer aggressiveness and progression. *Oncotarget* 2016;7(49):81156-71. doi: 10.18632/oncotarget.13215
30. Steele EM, Holmes JA, editors. A review of salvage treatment options for disease progression after radiation therapy for localized prostate cancer. *Urologic Oncology: Seminars and Original Investigations*; 2019: Elsevier.
31. Tisseverasinghe SA, Crook JM. The role of salvage brachytherapy for local relapse after external beam radiotherapy for prostate cancer. *Transl Androl Urol* 2018;7(3):414-35. doi: 10.21037/tau.2018.05.09
32. Artibani W, Porcaro AB, De Marco V, Cerruto MA, Siracusano S. Management of biochemical recurrence after primary curative treatment for prostate cancer: A review. *Urol Int* 2018;100(3):251-62. doi: 10.1159/000481438
33. Wang C, Chen Q, Xu H. Wnt/beta-catenin signal transduction pathway in prostate cancer and associated drug resistance. *Discov Oncol* 2021;12(1):40. doi: 10.1007/s12672-021-00433-6
34. Wang X, Jiang B, Zhang Y. Gankyrin regulates cell signaling network. *Tumour Biol* 2016;37(5):5675-82. doi: 10.1007/s13277-016-4854-z
35. Zhang Y, Wang X. Targeting the wnt/beta-catenin signaling pathway in cancer. *J Hematol Oncol* 2020;13(1):165. doi: 10.1186/s13045-020-00990-3
36. Zhao X, Fu J, Xu A, Yu L, Zhu J, Dai R, et al. Gankyrin drives malignant transformation of chronic liver damage-mediated fibrosis via the rac1/jnk pathway. *Cell Death Dis* 2015;6(5):e1751. doi: 10.1038/cddis.2015.120
37. Moussavi M, Haddad F, Rassouli FB, Iranshahi M, Soleymanifard S. Synergy between auraptene, ionizing radiation, and anticancer drugs in colon adenocarcinoma cells. *Phytother Res* 2017;31(9):1369-75. doi: 10.1002/ptr.5863
38. Kypta RM, Waxman J. Wnt/beta-catenin signalling in prostate cancer. *Nat Rev Urol* 2012;9(8):418-28. doi: 10.1038/nrurol.2012.116